Role of mitochondrial ROS in the brain: from physiology to neurodegeneration

Plamena R. Angelova, Andrey Y. Abramov

Department of Molecular Neuroscience, UCL Institute of Neurology, Queen Square, London WC1N 3BG, UK

Correspondence to: Plamena R. Angelova p.stroh@ucl.ac.uk or Andrey Y. Abramov a.abramov@ucl.ac.uk

Abstract

Mitochondria are the key organellae in the cell which are responsible for the energy production, and control many processes from signalling to cell death. The function of the mitochondrial electron transport chain and several enzymes in the matrix and on the outer membrane of mitochondria is coupled with the production of reactive oxygen species in the form of superoxide anion or hydrogen peroxide. Considering the constant production of ROS species, mitochondria are protected by several highly efficient antioxidant systems. The rate of ROS production in mitochondria in the brain is variable depending on the availability of substrates, the partial pressure of oxygen and on many other factors. Such rapidly changing levels of reactive oxygen species in mitochondria, coupled with multiple essential cellular functions, render ROS to be adapted in physiological signalling. Thus any mutations, environmental toxins and chronic ischaemic conditions could affect the mitochondrial redox balance and lead to the development of pathology. In long-living and non-mitotic cells like neurons, oxidative stress induced by overproduction of mitochondrial ROS or impairment of the antioxidant defence results in a dysfunction of mitochondria and initiation of a cell death cascade. Mitochondrial ROS overproduction and changes in mitochondrial redox homeostasis has been shown to be involved in a number of neurological conditions and in greater part of the neurodegenerative diseases. Here, we summarise the involvement of mitochondrial ROS in the mechanism of neuronal loss of major neurodegenerative disorders.

Introduction

Mitochondria are organellae which play multiple important functions in the cell. Despite versatility of the duties, the main function of mitochondria is producing energy in form of ATP and almost all other processes inside of mitochondria are connected or dependent on bioenergetics. ATP synthesis by oxidative phosphorylation is coupled with mitochondrial respiration. Respiration is generating mitochondrial transmembrane potential by pumping the protons by mitochondrial complexes I, III and IV of electron transport chain (ETC). Mitochondrial membrane potential $\Delta\psi m$ is a cross linked element in mitochondrial function – it used as a proton motive force for ATP synthesis, help to maintain the shape of this organelle and keep mitochondrial pro-apoptotic proteins, which released to cytosol in case of $\Delta\psi m$ collapse.

Distribution of mitochondrial in the cells from diverse tissues is dependent on the energy demand. However, despite the density of mitochondria in myocytes is higher than in neurons, brain consumes almost 10 times more oxygen and glucose then other tissues. Considering high energy demand and high rate of ATP production and consumption in the brain most of mitochondrial mutations or mitochondrial toxins damage brain function and lead to neurological pathology [1].

Mitochondria produce free radicals, which due to high accessibility of the oxygen in this organelle are mostly reactive oxygen species (ROS). Although mitochondria produce ROS in number of enzymes, the vast majority of the free radicals which named in the literature "mitochondrial ROS" are produced in ETC. The rate of ROS production, mitochondrial membrane potential ($\Delta\psi m$) and the activity of the complexes of the ETC are highly interdependent [2]. Therefore dissipation of the mitochondrial membrane potential on one hand could lead to an increase in ROS generation when respiration is inhibited. On the other hand, if drop in the $\Delta\psi m$ is stimulated by uncoupling, it could lead to a reduced rate of free radical production. Similarly, hyperpolarisation of mitochondria could lead to an increase of ROS production. Thus, production of ROS in the ETC is dependent on the release of electrons out of the electron transport chain followed by the formation of free radicals. The process of release of electrons could be induced by reverse flux of electrons and activity of complex I and II as a donor of electrons and partial inhibition of the complexes by hyperpolarisation, ischaemic conditions or by chemical compounds [3;4] (Figure 1). Although hydrogen peroxide was a first ROS which was shown to be produced in mitochondria [5;6];[7], electron escape from ETC generate free radical predominantly in the form of super oxide radical $O_2^{\cdot-}$ which later converts to H_2O_2.

Despite the fact that respiratory chain is a major ROS producer in mitochondria under resting conditions, several matrix proteins and complexes, including enzymes of TCA cycle such an aconitase, pyruvate dehydrogenase (PDH), and α-ketoglutarate dehydrogenase (αKGDH) could produce $O_2^{\cdot-}$ [8];[9]. Some inner mitochondrial membrane proteins (various cytochrome P450 enzymes, glycerol-3-phosphate dehydrogenase) which activity is partially dependent on $\Delta\psi m$ are also could produce ROS. Located on the outer membrane of mitochondria, enzyme monoamine oxidase (MAO) is utilizing monoamines producing aldehydes and hydrogen peroxide, another outer membrane protein cytochrome b5 reductase (Cb5R) which also can produce ROS (Figure 1).

Redox balance in the mitochondrial matrix is maintained by effective antioxidant system. Lifetime of the superoxide anion is 1 nanosecond and it can rapidly dismutate spontaneously or enzymatically with the help of manganese superoxide dismutase in mitochondrial matrix or by Cu, Zn-superoxide dismutase in the intermembrane space to hydrogen peroxide. Major endogenous antioxidant glutathione (GSH) is distributed through mitochondria and the rest of the cell and isolate major peptides from oxidation by $O_2^{\cdot-}$ or other forms of ROS. The permeable H_2O_2 is participating

in signalling cascades and degraded by enzymes catalase (CAT), glutathione peroxidase (GPx), and peroxiredoxin 3 [10]; [11].

One of the major initial form of ROS in mitochondria is superoxide anion radical $O_2^{\cdot-}$. Considering very short lifetime of this free radical ($\sim 1*10^{-9}$ second) it is very unlikely that superoxide can play role in physiology but possibly it can induce oxidative damage in neighbouring area. Superoxide dismutase (SOD1) is more likely play signalling rather antioxidative enzyme because it convert $O_2^{\cdot-}$ to more stable hydrogen peroxide which can be transported as signalling molecule (Figure 1). However, H_2O_2 can become dangerous for cells when it produce most toxic form of ROS – hydroxyl anion in Fenton reaction [12].

Mitochondria possess a number of "tools" to produce ROS in response to extracellular (such as the decrease of the oxygen level, toxins, increase of glucose uptake), cellular (hormones, transmitters), or intramitochondrial (availability of substrates) triggers. Mitochondrially generated ROS (from MAO) can stimulate lipid peroxidation which activates phospholipase C and IP3-triggered calcium signal [13]; [14]; [15] (Figure 1). Mitochondrial ROS increase in response to hypoxia stimulate calcium signal in astrocytes [16] and actives respiration [17]. Mitochondrial calcium uptake is redox sensible and can be regulated by ROS [18]. More prolonged elevation of ROS in mitochondria shown to be involved in the number of cell processes including proliferation of the cells [19]. However, any well balanced systems, even such a mitochondria, could be disrupted that result in pathology. Overproduction of ROS or dysregulation of antioxidant system lead to a number of pathologies, in brain it leads to cell death and neurodegeneration.

Neurodegenerative diseases are progressive, devastating and incurable, and are becoming increasingly prevalent in our aging populations. Aging of the population around world brings the neurodegenerative diseases to be one of the top medical and social problems. Thus, two major neurodegenerative disorders – Alzheimer's disease (AD) and Parkinson's Disease (PD) affecting 5% (AD) and 1 % (PD) of individuals aged 65 and above [20]; [21]. The annual cost in nursing home care for major neurodegenerative disorders in European countries is estimated to be in hundreds of million euros [22].

Neurodegenerative diseases, including Alzheimer's disease, Parkinson's disease, motor neuron disease, and Huntington disease all share a several common feature such an accumulation of abnormally aggregated proteins termed pathological inclusions and involvement of oxidative damage and mitochondrial dysfunction in pathogenesis. Many of the genes associated with PD, ALS or ataxias are linked to mitochondria. All aggregated misfolded proteins which involved in neurodegenerative disorders (β-amyloid, tau, α-synuclein and huntingtin) inhibiting mitochondrial function and induce oxidative stress [23;24]. Importantly, mutations in mitochondrial DNA result not only in mitochondrial myopathy, encephalopathy, lactic acidosis, and stroke-like episodes (MELAS), or myoclonic epilepsy with ragged red fibers (MERRF) but also to PD. Involvement of oxidative stress in the mechanism neuronal loss shown for the majority of neurodegenerative disorders [25]. However, antioxidant therapy approaches have failed as a treatment on a clinical level for the most of these diseases.

Although mitochondria produced far less ROS compare to NADPH oxidase, in long-lived neurons, where active mitochondria function must be maintained for an entire lifetime, implication of mitochondrial ROS in physiology and pathology may be crucial. Here we review the role of mitochondrial ROS in pathology of neurodegeneration.

Alzheimer's disease. Alzheimer's disease (AD) is a most common neurodegenerative disorder affecting aged population. Pathology of AD is characterised with senile plagues (consists predominantly with aggregated β-amyloid) and intracellular neurofibrillary tangles (formed by tau aggregates). The involvement of mitochondria in the mechanism

of AD pathology is not so direct, when compared to other neurodegenerative disorders, but the role of oxidative stress and mitochondrial dysfunction is shown for diverse models of AD [24]. Thus, reduction in complex IV activity has been demonstrated in mitochondria from the hippocampus and platelets of AD patients and in AD cybrid cells [26;27] [1]. Aggregation of βA leads to oxidative stress, mitochondrial dysfunction and energy failure prior to the development of plaque pathology [28].

Aggregated βA can reduce of mitochondrial respiration in neurons and astrocytes through the inhibition of complexes I and IV [29];[30]. Inhibition of ETC potentially can induce ROS production but direct increase in mitochondrial ROS production was shown in relatively small amount of publication [31];[32]. However, superoxide production from mitochondria but not from NADPH oxidase shown to be associated with blocked long-term potentiation in Tg2576 mouse model of AD [33]. Importance of mitochondrial ROS was also confirmed by results with mitochondrially located antioxidant MitoQ, which prevented cognitive decline, Aβ accumulation, astrogliosis, and synaptic loss in triple transgenic mouse model of AD [34]. MitoQ extends lifespan and improves health span of a transgenic Caenorhabditis elegans model of Alzheimer disease [35]. The role of oxidative damage in sporadic AD is confirmed in experiments where inhibition of lipid peroxidation with deuterium-reinforced polyunsaturated fatty acids diet improve cognition and memory in in aldehyde dehydrogenase 2 (Aldh2) null mice, an established model of oxidative stress-related cognitive impairment that exhibits AD-like pathologies [36]. Olfactory bulbectomy lead to the strong AD phenotype in mice. In mice, neurodegeneration which caused by olfactory bulbectomy is accompanied by energy metabolism disturbances and oxidative stress in brain mitochondria similar to those occurring in transgenic animals - familial AD models and patients with sporadic AD [37].

Many cases of autosomal dominant early onset Alzheimer's disease (AD) result from mutations in the genes encoding in presenilins (PS1 or PS2). Mitochondrial ROS shown to be important in the mechanism of the triggering of mitochondrial permeability transition pore (PTP) and activation process of cell death in PS1 cells [38].

Mitochondrial ROS production in AD models is much smaller comparing to effects on the production of ROS in NADPH oxidase [39] [40]. However, production of ROS in NADPH oxidase lead to mitochondrial depolarisation due to lack of substrates due activation of the DNA-repairing enzyme PARP [41]; [42]; [43]. Combination of calcium and ROS production under β-amyloid stimulation induce opening mitochondrial PTP and cell death [44]; [45]; [46]. Prevention of PTP opening by inducing cyclophilin D deficiency (molecular blocker of PTP opening) is also able to improve mitochondrial function and learning/memory in an aging Alzheimer disease mouse model [47].

Oxidative stress is proven to be one of the major trigger for pathology in AD [1] but mitochondrial is shown to be a target for oxidative damage rather than source of ROS production.

Familial form of frontotemporal dementia is induced by mutation in MAPT gene, encoding tau. Function of mitochondria is altered in the neurons of these patients. It results in the higher mitochondrial membrane potential with overproduction of the ROS in mitochondria, which in turn causes oxidative stress and cell death. Mitochondrial ROS overproduction in these cells is a major trigger for neuronal cell death and can be prevented by mitochondrial antioxidants [48].

Vascular diseases causing dementia. Vascular dementia is the cognitive decline resulting from cerebral vasculature hypoperfusion. A chronic hypoperfusion or blockade of a brain blood vessel could lead to a damage of the surrounding brain tissue and build-up of toxic waste substances and could result in various conditions, e.g. cerebral small vessel disease (CSVD) induced by hypercholesterolemia, cerebral amyloid angiopathy (CAA), stroke and ischaemia reperfusion injury, etc. Very often vascular dementia is a prerequisite for the development of Alzheimer's

disease when a defect clearance of β- amyloid is present. Undeniably, all these conditions have the same output phenotype - impairment of mitochondrial function and increased oxidative stress due to chronic hypoxia and substrate deprivation. Oxidative stress and mitochondrial dysfunction has been linked to the development of dementia in majority of the cases [97]. Supplementation with last generation antioxidants Resveratrol in form of solid lipid nanoparticles or Edaravone has been very promising. This type of treatment could activate Nrf-2/ HO-1 pathway and could mitigate the mitochondrial ROS production, the consequent lipid peroxidation, protein carbonyls formation and to improve the Mn-SOD activity in a permanent bilateral common carotid artery occlusion rodent model of vascular dementia [98;99].

Parkinson's Disease. There is much evidence that oxidative stress occurs in, and contributes to, the pathogenesis of PD. Post mortem studies of the brains of patients with PD reveal increased levels of lipid peroxidation markers (malondialdehyde and 4-hydroxynonenal) and the presence of protein oxidative damage in the form of protein carbonyls [49]. It has been reported that there is an increase in the common deletions in mtDNA in the surviving dopaminergic neurons in the substantia nigra of patients with PD. These deletions are caused by oxidative stress [50]. Both the toxin and genetic models of PD also demonstrate increased oxidative stress which is connected with mitochondrial function. Inhibitors of mitochondrial complex I, rotenone or 1-methyl-4-phenylpyridinium (MPP+) produces superoxide anions in submitochondrial particles, and the neurotoxic effects of MPP+ and rotenone are probably caused by oxidative stress rather than metabolic changes because they can be can be effectively prevented by treatment with antioxidants [51]. Importantly, mutations in mitochondrial complex I lead to neurodegeneration. In neurons derived from stem cell cybrids that contain such mtDNA mutations, the major trigger for cell death is also overproduction of superoxide in the matrix of mitochondria, but not an energy deprivation [52]. Mild uncoupling of mitochondria with mitochondrial uncoupling protein-2 (UCP-2) overexpression reduces ROS production in toxic (MPP+, rotenone) mouse model of PD. UCP-2 deficiency also increases the sensitivity of dopamine neurons to MPTP, whereas UCP-2 overexpression decreases MPTP-induced nigral dopamine cell loss [53]. Mutations in Parkinson protein-1 (PARK-1, which is also known as DJ-1) cause a rare autosomal-recessive form of PD. Loss of function of DJ-1 results in oxidative stress, and DJ-1 exerts neuroprotection via its antioxidant mechanism in mitochondria [54] [55]. DJ-1 knockout mice demonstrate increased mitochondrial oxidant stress and downregulation of mitochondrial uncoupling proteins [56].

Mutations in PINK1 cause a recessive form of PD. PINK1 is a mitochondrial kinase, and we and other authors have previously demonstrated that PINK1 deficiency results in impaired respiration with inhibition of complex I, and rotenone-like increased production of ROS in mitochondria [57]; [58]. Fibroblasts from patients with PINK1-associated PD exhibit impaired oxidative phosphorylation and oxidative stress [59]; [60]. Excessive ROS production in mitochondrial of PINK1 knockout neurons can be an inductor of inhibition of mitochondrial Na^{2+}/Ca^{2+} exchanger or glucose transporter and can be rescued by antioxidants [57]; [61]; [62] (Figure 1). Activation of Nrf2 by pharmacological activators restore mitochondrial metabolism in PINK1 deficient cells [63]. Additional ROS production in monoamine oxidase under application of dopamine induce opening of mitochondrial PTP and cell death in PINK1 deficient neurons [64].

It should be noted, that mitochondrial ROS play important role in pathology of PINK1 (mutation or deficiency) but also in physiology of PINK1/Parkin related mitophagy. Mitochondrial ROS production has been shown to be important for induction of mitochondrial recruitment of Parkin and initiation of mitophagy [65].

Excessive ROS production in mitochondria the familial and sporadic form of PD damage DNA that activates DNAA repairing enzyme PARP which induce energy deprivation in neurons due to NAD consumption [66]; [67] (Figure 1). Both familial and sporadic forms of PD are characterised by forming of Lewy body which consists from aggregated α-synuclein. While monomeric α-synuclein play physiological role in synaptic transduction and mitochondrial bioenergetics [68] [69], oligomeric peptide become toxic for cells [23]. Oligomeric α-synuclein is detected in mitochondria [70] where it inhibit complex I [71];[72]. Despite the fact that α-synuclein induced oxidative stress can be quenched by application coenzyme Q10 [73], the effect of any form of α-synuclein on mitochondrial ROS production was not identified [74]. Oligomeric α-synuclein is producing ROS independently of known enzymatic pathways that affect mitochondrial function and induced lipid peroxidation [75]; [74].

Progressive Supranuclear Palsy (PSP) is a form of atypical Parkinsonism which is characterised by accumulation of 4R tau inclusions and classified as tauopathy. The microtubule associated protein tau (MAPT) H1 haplotype is the major genetic risk factor associated with PSP, but recently many genes encoding proteins important in mitochondrial function or oxidative stress management, e.g. debrisoquine 4-hydroxylase (CYP2D6), paraoxonase (PON) 1 and 2, N-acetyltransferase (NAT) 1 and 2, and superoxide dismutase (SOD) 1 and 2 have also been implicated [93]. This links to findings that mitochondrial dysfunction and excess mitochondrial ROS production, lipid peroxidation as early as it occurs in mesenchymal stem cells from patients with the sporadic form of PSP pointing towards essential contribution of the cellular pathology. Importantly, even in the early developmental status the MSCs exhibit metabolically dysfunctional mitochondria and this negatively influences their differentiation capacity [94] and thus obscures the hope for autologous transplantation of mitochondria as a possible therapeutic direction for this disease.

Amyotrophic Lateral Sclerosis (ALS). The role of mitochondrially driven oxidative stress is linked to familial form of ALS with mutation in mitochondrial superoxide dismutase (SOD1). Amyotrophic lateral sclerosis (ALS) is a devastating neurodegenerative disease in which loss of spinal cord and cortical motor neurons starts leads to progressive paralysis and premature death [76];[77]. Mitochondrial oxidative damage has been demonstrated in patients affected by sporadic ALS [78]; [79] and in transgenic mice expressing a familial ALS-linked mutant Cu,Zn superoxide dismutase (SOD1) [80]. Importantly, reduction of the mitochondrial ROS in neurons with SOD1 mutant mouse model by generating double transgenic model with UCP2 (uncoupling protein 2) did not recover mitochondrial function and accelerates disease progression [81]. Mutations in RNA Transactivation response DNA-binding protein 43 (TDP-43), FUS/TLS and p62 are also associated with ALS and cells with these mutations also shown to have increase mitochondrial ROS and oxidative stress [82] [83].

Neurodegeneration with brain iron accumulation (NBIA). NBIA comprises of a heterogeneous group of diseases that a characterised by accumulation of iron in the basal ganglia and show mutation in PANK2 (pantothenate kinase). Pantothenate kinase (PANK2) mutation leads to a deficiency in CoA which in turn impairs energy metabolism in mitochondria. Acetyl CoA plays a role in synthesis and oxidation of fatty acids and oxidation of pyruvate in TCA. Animal models of the disease have failed so far because the rodent PANK-/- phenotype is not the typical neurodegenerative phenotype with brain iron accumulation and defective movements unless it is subjected to a ketogenic diet [85], the reason probably being the localisation of the murine PANK2 homolog is cytosolic in contrary to the human PANK2 that has been attributed to the mitochondria. Induced pluripotent stem cells (iPSC)-derived from PANK2 patient fibroblasts have been recently attempted to model the disease. These neurons have been shown to exhibit reduced glutathione levels and increased cytosolic and mitochondrial ROS production. Moreover,

supplementation with CoA has been shown to be protective [86]. Similarly, in our PANK2 iPSC-based model defective mitochondrial Complexes I and II function, increased ROS production and lower levels of cellular GSH which further resulted in increased lipid peroxidation have been demonstrated [87]. Application with the iron chelator Desferal further increased ROS production and further exacerbate the PANK2 phenotype.

However, mutations in several genes have been known to cause neurodegeneration with brain iron accumulation, e.g. PLA2G6, C19orf12, COASY, FA2H, ATP13A2, FTL/FTL1 etc. [84] and importantly, most of them are connected to oxidative stress.

PLA2G6 mutation is an autosomal recessive mutation in the gene encoding the calcium-independent phospholipase A2 located to the chromosome 22q12-q13 leads to infantile neuroaxonal dystrophy (INAD). Recently it has been discovered that PLA2G6 mutation lead as well to neurodegeneration with brain iron accumulation (NBIA) [88]. It has been reported that PLA2G6 mutation leads to development of early onset Parkinsonism [89]. Earlier on, a discovery that links the pathology of neurodegeneration with brain iron accumulation phospholipid metabolism to the disruption brain iron homeostasis has been shown [88;89]. Mitochondrial dysfunction, increased mitochondrial ROS generation and lipid peroxidation in fibroblasts from patients with PLA2G6 mutation was reported. Importantly, feeding Drosophila iPLA2-VIA-/- flies with deuterated PUFAs reduced the rates of lipids peroxidation to basal as well as partially rescue their locomotor deficits [90].

Friedrich's ataxia (FRDA). Cerebellar ataxia is caused by a mutation in the FXN gene, which leads to a GAA repeat expansion and lower availability of the protein frataxin, a key component for the formation of the Fe-S clusters of the mitochondrial complexes I, II and III from the electron transport chain (ETC). Increased levels of mitochondrial (and cytosolic) ROS production and level of lipid peroxidation from cerebellar granule cells from FRDA granule cells [91] is due to a inhibition of the mitochondrial respiration complex I and abnormal accumulation of iron in the mitochondria. In fibroblasts from two FRDA mouse models, YG8R and KIKO, inhibition of lipid peroxidation with deuterated PUFAs and Nrf-2 activators, TBE31 and Sulforaphane, could prevent the lipid peroxidation damage and the consequent cell death in these cells [92].

Huntington's disease (HD). HD-an autosomal dominant mutation of the mhtt gene, arises as a result a CAG repeat expansion of the gene coding the protein huntingtin. Similar to many other neurodegenerative diseases, oxidative stress and inflammation are heavily implicated. Characteristic for the HD brain samples are the increased levels of superoxide dismutase (Zn/Cu-SOD and mitochondrial Mn-SOD), glutathione peroxidases (GPx) and catalase (CAT), but not any canonical antioxidant response elements (AREs) gene products (NQO1, GCLM, GCLC, HMOX1/HO-1) [95]. However, neither overexpression of cytosolic Zn/Cu-SOD or mitochondrial Mn-SOD nor nutritional supplementation with α-tocopherol and coenzyme Q10 has led to prolongation of the lifespan of Drosophila HD model flies. In contrary, activation of the Nrf2 pathway by SIRT2 inhibition and induction of NQO1 has been very [promising and effective in protecting oxidative damage in rodent and human HD models [96].

Conclusions

Mitochondria extensively generate ROS or/and are targeted by free radicals in the aetiopathology of the major neurodegenerative diseases. In most of the diseases overproduction of ROS or loss of function of antioxidant pathways lead to oxidative damage of biological molecules. This in turn leads to a deregulation in the function they are responsible for or ultimately to the initiation of cell death. Thus, even small increase in ROS production over the basal rates requires elevated antioxidant activity. Maintenance of the major antioxidant systems (in brain predominantly

GSH) is a highly energy consuming process and any increased activity of antioxidants production may lead to limitation of substrates for the normal functioning of mitochondria (Figure 2).

Mitochondrial redox balance and physiological role of mitochondrial ROS are very important in neuronal housekeeping. And despite the vast amount of publications which confirm damaging role of mitochondrial ROS in neurodegeneration, mitochondrially targeted antioxidants are not effective in treatment of neurodegenerative diseases on a patient level due various reasons, including quenching effect on the physiological signalling function of ROS. Antioxidant therapy has been proven to be effective for a number of neurodegenerative disorders in experiments on a cellular level, but most of the clinical trials have failed to demonstrate neuroprotection or efficacy in patients. This failure to translate the positive effects of antioxidants is usually attributed to the difficulties in antioxidant delivery to cells in the brain, or by the chemical instability of antioxidants. We propose that oxidative damage in neurodegeneration should be prevented or restricted through the direct inhibition of ROS production from specific sources, rather than through the use of scavengers. Furthermore finding the ways to quench the production of free radicals in these cells specifically, either through direct inhibition of an enzyme, or by increasing the endogenous antioxidants or by increasing energy production, represents one of the most promising directions of development for therapeutic strategies.

Figure Legends

Figure 1. Mitochondria is a producer and target of ROS. Mitochondria generating ROS in ETC, TCA cycle enzymes and monoamine oxidase (MAO). Production of hydrogens peroxide in MAO, or superoxide in ETC can stimulate lipid peroxidation and IP3 dependent calcium signal. Overproduction of ROS in NADPH oxidase in Parkinson's disease or Alzheimer's disease activates PARP which consume NAD and reduce NADH for complex I. Mitochondrial ROS production can inhibit glucose transporter and induce limitation of mitochondrial substrates for mitochondria.

Figure 2. Balance and interrelation between ROS production, energy metabolism and antioxidant homeostasis. Any changes in this balance lead to oxidative damage.

References

[1] Gandhi S & Abramov AY (2012) Mechanism of oxidative stress in neurodegeneration. *Oxid. Med. Cell Longev.*, **2012,** 428010.

[2] Votyakova TV & Reynolds IJ (2001) DeltaPsi(m)-Dependent and -independent production of reactive oxygen species by rat brain mitochondria. *J. Neurochem.*, **79,** 266-277.

[3] Vinogradov AD & Grivennikova VG (2016) Oxidation of NADH and ROS production by respiratory complex I. *Biochim. Biophys. Acta*, **1857,** 863-871.

[4] Kovac S, Angelova PR, Holmstrom KM, Zhang Y, Dinkova-Kostova AT, & Abramov AY (2015) Nrf2 regulates ROS production by mitochondria and NADPH oxidase. *Biochim. Biophys. Acta*, **1850,** 794-801.

[5] Loschen G, Flohe L, & Chance B (1971) Respiratory chain linked H(2)O(2) production in pigeon heart mitochondria. *FEBS Lett.*, **18,** 261-264.

[6]   Boveris A, Oshino N, & Chance B (1972) The cellular production of hydrogen peroxide. *Biochem. J.*, **128,** 617-630.

[7]   Loschen G, Azzi A, Richter C, & Flohe L (1974) Superoxide radicals as precursors of mitochondrial hydrogen peroxide. *FEBS Lett.*, **42,** 68-72.

[8]   Gibson GE, Blass JP, Beal MF, & Bunik V (2005) The alpha-ketoglutarate-dehydrogenase complex: a mediator between mitochondria and oxidative stress in neurodegeneration. *Mol. Neurobiol.*, **31,** 43-63.

[9]   Starkov AA, Fiskum G, Chinopoulos C, Lorenzo BJ, Browne SE, Patel MS, & Beal MF (2004) Mitochondrial alpha-ketoglutarate dehydrogenase complex generates reactive oxygen species. *J. Neurosci.*, **24,** 7779-7788.

[10] Forman HJ, Maiorino M, & Ursini F (2010) Signaling functions of reactive oxygen species. *Biochemistry*, **49,** 835-842.

[11] Mukherjee SP & Mukherjee C (1982) Similar activities of nerve growth factor and its homologue proinsulin in intracellular hydrogen peroxide production and metabolism in adipocytes. Transmembrane signalling relative to insulin-mimicking cellular effects. *Biochem. Pharmacol.*, **31,** 3163-3172.

[12] Wardman P & Candeias LP (1996) Fenton chemistry: an introduction. *Radiat. Res.*, **145,** 523-531.

[13] Vaarmann A, Gandhi S, & Abramov AY (2010) Dopamine induces Ca2+ signaling in astrocytes through reactive oxygen species generated by monoamine oxidase. *J. Biol. Chem.*, **285,** 25018-25023.

[14] Domijan AM, Kovac S, & Abramov AY (2014) Lipid peroxidation is essential for phospholipase C activity and the inositol-trisphosphate-related Ca(2)(+) signal. *J. Cell Sci.*, **127,** 21-26.

[15] Angelova PR & Abramov AY (2016) Functional role of mitochondrial reactive oxygen species in physiology. *Free Radic. Biol. Med.*.

[16] Abramov AY, Scorziello A, & Duchen MR (2007) Three distinct mechanisms generate oxygen free radicals in neurons and contribute to cell death during anoxia and reoxygenation. *J. Neurosci.*, **27,** 1129-1138.

[17] Angelova PR, Kasymov V, Christie I, Sheikhbahaei S, Turovsky E, Marina N, Korsak A, Zwicker J, Teschemacher AG, Ackland GL, Funk GD, Kasparov S, Abramov AY, & Gourine AV (2015) Functional Oxygen Sensitivity of Astrocytes. *J. Neurosci.*, **35,** 10460-10473.

[18] Dong Z, Shanmughapriya S, Tomar D, Siddiqui N, Lynch S, Nemani N, Breves SL, Zhang X, Tripathi A, Palaniappan P, Riitano MF, Worth AM, Seelam A, Carvalho E, Subbiah R, Jana F, Soboloff J, Peng Y, Cheung JY, Joseph SK, Caplan J, Rajan S, Stathopulos PB, & Madesh M (2017) Mitochondrial Ca2+ Uniporter Is a Mitochondrial Luminal Redox Sensor that Augments MCU Channel Activity. *Mol. Cell*, **65,** 1014-1028.

[19] Diebold L & Chandel NS (2016) Mitochondrial ROS regulation of proliferating cells. *Free Radic. Biol. Med.*, **100,** 86-93.

[20] Bras J, Singleton A, Cookson MR, & Hardy J (2008) Emerging pathways in genetic Parkinson's disease: Potential role of ceramide metabolism in Lewy body disease. *FEBS J.*, **275,** 5767-5773.

[21] Hardy J (2017) The discovery of Alzheimer-causing mutations in the APP gene and the formulation of the "amyloid cascade hypothesis". *FEBS J.*, **284,** 1040-1044.

[22] Vossius C, Nilsen OB, & Larsen JP (2009) Parkinson's disease and nursing home placement: the economic impact of the need for care. *Eur. J. Neurol.*, **16,** 194-200.

[23] Angelova PR & Abramov AY (2017) Alpha-synuclein and beta-amyloid - different targets, same players: calcium, free radicals and mitochondria in the mechanism of neurodegeneration. *Biochem. Biophys. Res. Commun.*, **483,** 1110-1115.

[24] Abramov AY, Berezhnov AV, Fedotova EI, Zinchenko VP, & Dolgacheva LP (2017) Interaction of misfolded proteins and mitochondria in neurodegenerative disorders. *Biochem. Soc. Trans.*.

[25] Gandhi S & Abramov AY (2012) Mechanism of oxidative stress in neurodegeneration. *Oxid. Med. Cell Longev.*, **2012,** 428010.

[26] Sheehan JP, Swerdlow RH, Miller SW, Davis RE, Parks JK, Parker WD, & Tuttle JB (1997) Calcium homeostasis and reactive oxygen species production in cells transformed by mitochondria from individuals with sporadic Alzheimer's disease. *J. Neurosci.*, **17,** 4612-4622.

[27] Du H, Guo L, Yan S, Sosunov AA, McKhann GM, & Yan SS (2010) Early deficits in synaptic mitochondria in an Alzheimer's disease mouse model. *Proc. Natl. Acad. Sci. U. S. A*, **107,** 18670-18675.

[28] Caspersen C, Wang N, Yao J, Sosunov A, Chen X, Lustbader JW, Xu HW, Stern D, McKhann G, & Yan SD (2005) Mitochondrial Abeta: a potential focal point for neuronal metabolic dysfunction in Alzheimer's disease. *FASEB J.*, **19,** 2040-2041.

[29] Casley CS, Canevari L, Land JM, Clark JB, & Sharpe MA (2002) Beta-amyloid inhibits integrated mitochondrial respiration and key enzyme activities. *J. Neurochem.*, **80,** 91-100.

[30] Canevari L, Clark JB, & Bates TE (1999) beta-Amyloid fragment 25-35 selectively decreases complex IV activity in isolated mitochondria. *FEBS Lett.*, **457,** 131-134.

[31] Yu F, Gong P, Hu Z, Qiu Y, Cui Y, Gao X, Chen H, & Li J (2015) Cu(II) enhances the effect of Alzheimer's amyloid-beta peptide on microglial activation. *J. Neuroinflammation.*, **12,** 122.

[32] Aminzadeh M, Roghani M, Sarfallah A, & Riazi GH (2017) TRPM2 dependence of ROS-induced NLRP3 activation in Alzheimer's disease. *Int. Immunopharmacol.*, **54,** 78-85.

[33] Ma T, Hoeffer CA, Wong H, Massaad CA, Zhou P, Iadecola C, Murphy MP, Pautler RG, & Klann E (2011) Amyloid beta-induced impairments in hippocampal synaptic plasticity are rescued by decreasing mitochondrial superoxide. *J. Neurosci.*, **31,** 5589-5595.

[34] McManus MJ, Murphy MP, & Franklin JL (2011) The mitochondria-targeted antioxidant MitoQ prevents loss of spatial memory retention and early neuropathology in a transgenic mouse model of Alzheimer's disease. *J. Neurosci.*, **31,** 15703-15715.

[35] Ng LF, Gruber J, Cheah IK, Goo CK, Cheong WF, Shui G, Sit KP, Wenk MR, & Halliwell B (2014) The mitochondria-targeted antioxidant MitoQ extends lifespan and improves healthspan of a transgenic Caenorhabditis elegans model of Alzheimer disease. *Free Radic. Biol. Med.*, **71,** 390-401.

[36] Elharram A, Czegledy NM, Golod M, Milne GL, Pollock E, Bennett BM, & Shchepinov MS (2017) Deuterium-reinforced polyunsaturated fatty acids improve cognition in a mouse model of sporadic Alzheimer's disease. *FEBS J.*, **284,** 4083-4095.

[37] Avetisyan AV, Samokhin AN, Alexandrova IY, Zinovkin RA, Simonyan RA, & Bobkova NV (2016) Mitochondrial Dysfunction in Neocortex and Hippocampus of Olfactory Bulbectomized Mice, a Model of Alzheimer's Disease. *Biochemistry (Mosc.)*, **81,** 615-623.

[38] Keller JN, Guo Q, Holtsberg FW, Bruce-Keller AJ, & Mattson MP (1998) Increased sensitivity to mitochondrial toxin-induced apoptosis in neural cells expressing mutant presenilin-1 is linked to perturbed calcium homeostasis and enhanced oxyradical production. *J. Neurosci.*, **18,** 4439-4450.

[39] Abramov AY & Duchen MR (2005) The role of an astrocytic NADPH oxidase in the neurotoxicity of amyloid beta peptides. *Philos. Trans. R. Soc. Lond B Biol. Sci.*, **360,** 2309-2314.

[40] Hashimoto Y, Kaneko Y, Tsukamoto E, Frankowski H, Kouyama K, Kita Y, Niikura T, Aiso S, Bredesen DE, Matsuoka M, & Nishimoto I (2004) Molecular characterization of neurohybrid cell death induced by Alzheimer's amyloid-beta peptides via p75NTR/PLAIDD. *J. Neurochem.*, **90,** 549-558.

[41] Abramov AY, Canevari L, & Duchen MR (2004) Beta-amyloid peptides induce mitochondrial dysfunction and oxidative stress in astrocytes and death of neurons through activation of NADPH oxidase. *J. Neurosci.*, **24,** 565-575.

[42] Abeti R, Abramov AY, & Duchen MR (2011) Beta-amyloid activates PARP causing astrocytic metabolic failure and neuronal death. *Brain*, **134,** 1658-1672.

[43] Narayan P, Holmstrom KM, Kim DH, Whitcomb DJ, Wilson MR, St George-Hyslop P, Wood NW, Dobson CM, Cho K, Abramov AY, & Klenerman D (2014) Rare individual amyloid-beta oligomers act on astrocytes to initiate neuronal damage. *Biochemistry*, **53,** 2442-2453.

[44] Abramov AY, Canevari L, & Duchen MR (2004) Calcium signals induced by amyloid beta peptide and their consequences in neurons and astrocytes in culture. *Biochim. Biophys. Acta*, **1742,** 81-87.

[45] Abramov AY & Duchen MR (2005) The role of an astrocytic NADPH oxidase in the neurotoxicity of amyloid beta peptides. *Philos. Trans. R. Soc. Lond B Biol. Sci.*, **360,** 2309-2314.

[46] Shevtsova EF, Vinogradova DV, Neganova ME, Avila-Rodriguez M, Ashraf GM, Barreto GE, Bachurin SO, & Aliev G (2017) Mitochondrial Permeability Transition Pore as a Suitable Targ e t for Neuroprotective Agents Against Alzheimer's Disease. *CNS. Neurol. Disord. Drug Targets.*, **16,** 677-685.

[47] Du H, Guo L, Fang F, Chen D, Sosunov AA, McKhann GM, Yan Y, Wang C, Zhang H, Molkentin JD, Gunn-Moore FJ, Vonsattel JP, Arancio O, Chen JX, & Yan SD (2008) Cyclophilin D deficiency attenuates mitochondrial and neuronal perturbation and ameliorates learning and memory in Alzheimer's disease. *Nat. Med.*, **14,** 1097-1105.

[48] Esteras N, Rohrer JD, Hardy J, Wray S, & Abramov AY (2017) Mitochondrial hyperpolarization in iPSC-derived neurons from patients of FTDP-17 with 10+16 MAPT mutation leads to oxidative stress and neurodegeneration. *Redox. Biol.*, **12,** 410-422.

[49] Beal MF (2002) Oxidatively modified proteins in aging and disease. *Free Radic. Biol. Med.*, **32,** 797-803.

[50] Bender A, Krishnan KJ, Morris CM, Taylor GA, Reeve AK, Perry RH, Jaros E, Hersheson JS, Betts J, Klopstock T, Taylor RW, & Turnbull DM (2006) High levels of mitochondrial DNA deletions in substantia nigra neurons in aging and Parkinson disease. *Nat. Genet.*, **38,** 515-517.

[51] Gonzalez-Polo RA, Soler G, Rodriguezmartin A, Moran JM, & Fuentes JM (2004) Protection against MPP+ neurotoxicity in cerebellar granule cells by antioxidants. *Cell Biol. Int.*, **28,** 373-380.

[52] Abramov AY, Smulders-Srinivasan TK, Kirby DM, Acin-Perez R, Enriquez JA, Lightowlers RN, Duchen MR, & Turnbull DM (2010) Mechanism of neurodegeneration of neurons with mitochondrial DNA mutations. *Brain*, **133,** 797-807.

[53] Andrews ZB, Horvath B, Barnstable CJ, Elsworth J, Yang L, Beal MF, Roth RH, Matthews RT, & Horvath TL (2005) Uncoupling protein-2 is critical for nigral dopamine cell survival in a mouse model of Parkinson's disease. *J. Neurosci.*, **25,** 184-191.

[54] Lev N, Ickowicz D, Barhum Y, Melamed E, & Offen D (2009) DJ-1 changes in G93A-SOD1 transgenic mice: implications for oxidative stress in ALS. *J. Mol. Neurosci.*, **38,** 94-102.

[55] Li HM, Niki T, Taira T, Iguchi-Ariga SM, & Ariga H (2005) Association of DJ-1 with chaperones and enhanced association and colocalization with mitochondrial Hsp70 by oxidative stress. *Free Radic. Res.*, **39,** 1091-1099.

[56] Guzman JN, Sanchez-Padilla J, Wokosin D, Kondapalli J, Ilijic E, Schumacker PT, & Surmeier DJ (2010) Oxidant stress evoked by pacemaking in dopaminergic neurons is attenuated by DJ-1. *Nature*, **468,** 696-700.

[57] Gandhi S, Wood-Kaczmar A, Yao Z, Plun-Favreau H, Deas E, Klupsch K, Downward J, Latchman DS, Tabrizi SJ, Wood NW, Duchen MR, & Abramov AY (2009) PINK1-associated Parkinson's disease is caused by neuronal vulnerability to calcium-induced cell death. *Mol. Cell*, **33,** 627-638.

[58] Piccoli C, Sardanelli A, Scrima R, Ripoli M, Quarato G, D'Aprile A, Bellomo F, Scacco S, De MG, Filla A, Iuso A, Boffoli D, Capitanio N, & Papa S (2008) Mitochondrial respiratory dysfunction in familiar parkinsonism associated with PINK1 mutation. *Neurochem. Res.*, **33,** 2565-2574.

[59] Abramov AY, Gegg M, Grunewald A, Wood NW, Klein C, & Schapira AH (2011) Bioenergetic consequences of PINK1 mutations in Parkinson disease. *PLoS. One.*, **6,** e25622.

[60] Hoepken HH, Gispert S, Morales B, Wingerter O, Del TD, Mulsch A, Nussbaum RL, Muller K, Drose S, Brandt U, Deller T, Wirth B, Kudin AP, Kunz WS, & Auburger G (2007) Mitochondrial dysfunction, peroxidation damage and changes in glutathione metabolism in PARK6. *Neurobiol. Dis.*, **25,** 401-411.

[61] Kostic M, Ludtmann MH, Bading H, Hershfinkel M, Steer E, Chu CT, Abramov AY, & Sekler I (2015) PKA Phosphorylation of NCLX Reverses Mitochondrial Calcium Overload and Depolarization, Promoting Survival of PINK1-Deficient Dopaminergic Neurons. *Cell Rep.*.

[62] Deas E, Piipari K, Machhada A, Li A, Gutierrez-del-Arroyo A, Withers DJ, Wood NW, & Abramov AY (2014) PINK1 deficiency in beta-cells increases basal insulin secretion and improves glucose tolerance in mice. *Open. Biol.*, **4,** 140051.

[63] Dinkova-Kostova AT, Baird L, Holmstrom KM, Meyer CJ, & Abramov AY (2015) The spatiotemporal regulation of the Keap1-Nrf2 pathway and its importance in cellular bioenergetics. *Biochem. Soc. Trans.*, **43,** 602-610.

[64] Gandhi S, Vaarmann A, Yao Z, Duchen MR, Wood NW, & Abramov AY (2012) Dopamine induced neurodegeneration in a PINK1 model of Parkinson's disease. *PLoS. One.*, **7,** e37564.

[65] Xiao B, Goh JY, Xiao L, Xian H, Lim KL, & Liou YC (2017) Reactive oxygen species trigger Parkin/PINK1 pathway-dependent mitophagy by inducing mitochondrial recruitment of Parkin. *J. Biol. Chem.*, **292,** 16697-16708.

[66] Berger NA, Besson VC, Boulares AH, Burkle A, Chiarugi A, Clark RS, Curtin NJ, Cuzzocrea S, Dawson TM, Dawson VL, Hasko G, Liaudet L, Moroni F, Pacher P, Radermacher P, Salzman AL, Snyder SH, Soriano FG, Strosznajder RP, Sumegi B, Swanson RA, & Szabo C (2017) Opportunities for the repurposing of PARP inhibitors for the therapy of non-oncological diseases. *Br. J. Pharmacol.*.

[67] Delgado-Camprubi M, Esteras N, Soutar MP, Plun-Favreau H, & Abramov AY (2017) Deficiency of Parkinson's disease-related gene Fbxo7 is associated with impaired mitochondrial metabolism by PARP activation. *Cell Death. Differ.*, **24,** 120-131.

[68] Ludtmann MH, Angelova PR, Ninkina NN, Gandhi S, Buchman VL, & Abramov AY (2016) Monomeric Alpha-Synuclein Exerts a Physiological Role on Brain ATP Synthase. *J. Neurosci.*, **36,** 10510-10521.

[69] Burre J, Vivona S, Diao J, Sharma M, Brunger AT, & Sudhof TC (2013) Properties of native brain alpha-synuclein. *Nature*, **498,** E4-E6.

[70] Nakamura K, Nemani VM, Wallender EK, Kaehlcke K, Ott M, & Edwards RH (2008) Optical reporters for the conformation of alpha-synuclein reveal a specific interaction with mitochondria. *J. Neurosci.*, **28,** 12305-12317.

[71] Reeve AK, Ludtmann MH, Angelova PR, Simcox EM, Horrocks MH, Klenerman D, Gandhi S, Turnbull DM, & Abramov AY (2015) Aggregated alpha-synuclein and complex I deficiency: exploration of their relationship in differentiated neurons. *Cell Death. Dis.*, **6,** e1820.

[72] Luth ES, Stavrovskaya IG, Bartels T, Kristal BS, & Selkoe DJ (2014) Soluble, prefibrillar alpha-synuclein oligomers promote complex I-dependent, Ca2+-induced mitochondrial dysfunction. *J. Biol. Chem.*, **289,** 21490-21507.

[73] Esteves AR, Arduino DM, Swerdlow RH, Oliveira CR, & Cardoso SM (2009) Oxidative stress involvement in alpha-synuclein oligomerization in Parkinson's disease cybrids. *Antioxid. Redox Signal.*, **11,** 439-448.

[74] Deas E, Cremades N, Angelova PR, Ludtmann MH, Yao Z, Chen S, Horrocks MH, Banushi B, Little D, Devine MJ, Gissen P, Klenerman D, Dobson CM, Wood NW, Gandhi S, & Abramov AY (2016) Alpha-Synuclein Oligomers Interact with Metal Ions to Induce Oxidative Stress and Neuronal Death in Parkinson's Disease. *Antioxid. Redox Signal.*, **24,** 376-391.

[75] Angelova PR, Horrocks MH, Klenerman D, Gandhi S, Abramov AY, & Shchepinov MS (2015) Lipid peroxidation is essential for alpha-synuclein-induced cell death. *J. Neurochem.*, **133,** 582-589.

[76] Cozzolino M, Pesaresi MG, Gerbino V, Grosskreutz J, & Carri MT (2012) Amyotrophic lateral sclerosis: new insights into underlying molecular mechanisms and opportunities for therapeutic intervention. *Antioxid. Redox Signal.*, **17,** 1277-1330.

[77] Cozzolino M & Carri MT (2012) Mitochondrial dysfunction in ALS. *Prog. Neurobiol.*, **97,** 54-66.

[78] Shaw IC, Fitzmaurice PS, Mitchell JD, & Lynch PG (1995) Studies on cellular free radical protection mechanisms in the anterior horn from patients with amyotrophic lateral sclerosis. *Neurodegeneration.*, **4,** 391-396.

[79] Shibata N, Nagai R, Uchida K, Horiuchi S, Yamada S, Hirano A, Kawaguchi M, Yamamoto T, Sasaki S, & Kobayashi M (2001) Morphological evidence for lipid peroxidation and protein glycoxidation in spinal cords from sporadic amyotrophic lateral sclerosis patients. *Brain Res.*, **917,** 97-104.

[80] Shibata N (2001) Transgenic mouse model for familial amyotrophic lateral sclerosis with superoxide dismutase-1 mutation. *Neuropathology.*, **21,** 82-92.

[81] Peixoto PM, Kim HJ, Sider B, Starkov A, Horvath TL, & Manfredi G (2013) UCP2 overexpression worsens mitochondrial dysfunction and accelerates disease progression in a mouse model of amyotrophic lateral sclerosis. *Mol. Cell Neurosci.*, **57,** 104-110.

[82] Onesto E, Colombrita C, Gumina V, Borghi MO, Dusi S, Doretti A, Fagiolari G, Invernizzi F, Moggio M, Tiranti V, Silani V, & Ratti A (2016) Gene-specific mitochondria dysfunctions in human TARDBP and C9ORF72 fibroblasts. *Acta Neuropathol. Commun.*, **4,** 47.

[83] Bartolome F, Esteras N, Martin-Requero A, Boutoleau-Bretonniere C, Vercelletto M, Gabelle A, Le B, I, Honda T, Dinkova-Kostova AT, Hardy J, Carro E, & Abramov AY (2017) Pathogenic p62/SQSTM1 mutations impair energy metabolism through limitation of mitochondrial substrates. *Sci. Rep.*, **7,** 1666.

[84] Wiethoff S & Houlden H (2017) Neurodegeneration with brain iron accumulation. *Handb. Clin. Neurol.*, **145,** 157-166.

[85] Brunetti D, Dusi S, Morbin M, Uggetti A, Moda F, D'Amato I, Giordano C, d'Amati G, Cozzi A, Levi S, Hayflick S, & Tiranti V (2012) Pantothenate kinase-associated neurodegeneration: altered mitochondria membrane potential and defective respiration in Pank2 knock-out mouse model. *Hum. Mol. Genet.*, **21,** 5294-5305.

[86] Orellana DI, Santambrogio P, Rubio A, Yekhlef L, Cancellieri C, Dusi S, Giannelli SG, Venco P, Mazzara PG, Cozzi A, Ferrari M, Garavaglia B, Taverna S, Tiranti V, Broccoli V, & Levi S (2016) Coenzyme A corrects pathological defects in human neurons of PANK2-associated neurodegeneration. *EMBO Mol. Med.*, **8,** 1197-1211.

[87] Arber C, Angelova PR, Wiethoff S, Tsuchiya Y, Mazzacuva F, Preza E, Bhatia KP, Mills K, Gout I, Abramov AY, Hardy J, Duce JA, Houlden H, & Wray S (2017) iPSC-derived neuronal models of PANK2-associated neurodegeneration reveal mitochondrial dysfunction contributing to early disease. *PLoS. One.*, **12,** e0184104.

[88] Morgan NV, Westaway SK, Morton JE, Gregory A, Gissen P, Sonek S, Cangul H, Coryell J, Canham N, Nardocci N, Zorzi G, Pasha S, Rodriguez D, Desguerre I, Mubaidin A, Bertini E, Trembath RC, Simonati A, Schanen C, Johnson CA, Levinson B, Woods CG, Wilmot B, Kramer P, Gitschier J, Maher ER, & Hayflick SJ (2006) PLA2G6, encoding a phospholipase A2, is mutated in neurodegenerative disorders with high brain iron. *Nat. Genet.*, **38,** 752-754.

[89] Giri A, Guven G, Hanagasi H, Hauser AK, Erginul-Unaltuna N, Bilgic B, Gurvit H, Heutink P, Gasser T, Lohmann E, & Simon-Sanchez J (2016) PLA2G6 Mutations Related to Distinct Phenotypes: A New Case with Early-onset Parkinsonism. *Tremor. Other Hyperkinet. Mov (N. Y.)*, **6,** 363.

[90] Kinghorn KJ, Castillo-Quan JI, Bartolome F, Angelova PR, Li L, Pope S, Cocheme HM, Khan S, Asghari S, Bhatia KP, Hardy J, Abramov AY, & Partridge L (2015) Loss of PLA2G6 leads to elevated mitochondrial lipid peroxidation and mitochondrial dysfunction. *Brain*, **138,** 1801-1816.

[91] Abeti R, Parkinson MH, Hargreaves IP, Angelova PR, Sandi C, Pook MA, Giunti P, & Abramov AY (2016) 'Mitochondrial energy imbalance and lipid peroxidation cause cell death in Friedreich's ataxia'. *Cell Death. Dis.*, **7,** e2237.

[92] Abeti R, Uzun E, Renganathan I, Honda T, Pook MA, & Giunti P (2015) Targeting lipid peroxidation and mitochondrial imbalance in Friedreich's ataxia. *Pharmacol. Res.*, **99,** 344-350.

[93] Litvan I, Campbell G, Mangone CA, Verny M, McKee A, Chaudhuri KR, Jellinger K, Pearce RK, & D'Olhaberriague L (1997) Which clinical features differentiate progressive supranuclear palsy (Steele-Richardson-Olszewski syndrome) from related disorders? A clinicopathological study. *Brain*, **120 (Pt 1),** 65-74.

[94] Angelova PR, Barilani M, Lovejoy C, Dossena M, Vigano M, Seresini A, Piga D, Gandhi S, Pezzoli G, Abramov AY, & Lazzari L (2017) Mitochondrial dysfunction in Parkinsonian mesenchymal stem cells impairs differentiation. *Redox Biol.*, **14,** 474-484.

[95] Quinti L, Dayalan NS, Trager U, Chen X, Kegel-Gleason K, Lleres D, Connolly C, Chopra V, Low C, Moniot S, Sapp E, Tousley AR, Vodicka P, Van Kanegan MJ, Kaltenbach LS, Crawford LA, Fuszard M, Higgins M, Miller JRC, Farmer RE, Potluri V, Samajdar S, Meisel L, Zhang N, Snyder A, Stein R, Hersch SM, Ellerby LM, Weerapana E, Schwarzschild MA, Steegborn C, Leavitt BR, Degterev A, Tabrizi SJ, Lo DC, DiFiglia M, Thompson LM, Dinkova-Kostova AT, & Kazantsev AG (2017) KEAP1-modifying small molecule reveals muted NRF2 signaling responses in neural stem cells from Huntington's disease patients. *Proc. Natl. Acad. Sci. U. S. A*, **114,** E4676-E4685.

[96] Quinti L, Casale M, Moniot S, Pais TF, Van Kanegan MJ, Kaltenbach LS, Pallos J, Lim RG, Naidu SD, Runne H, Meisel L, Rauf NA, Leyfer D, Maxwell MM, Saiah E, Landers JE, Luthi-Carter R, Abagyan R, Dinkova-Kostova AT, Steegborn C, Marsh JL, Lo DC, Thompson LM, & Kazantsev AG (2016) S. *Cell Chem. Biol.*, **23,** 849-861.

[97] Liu H & Zhang J (2012) Cerebral hypoperfusion and cognitive impairment: the pathogenic role of vascular oxidative stress. *Int. J. Neurosci.*, **122,** 494-499.

[98] Yadav A, Sunkaria A, Singhal N, & Sandhir R (2017) Resveratrol loaded solid lipid nanoparticles attenuate mitochondrial oxidative stress in vascular dementia by activating Nrf2/HO-1 pathway. *Neurochem. Int.*.

[99] Zhang N, Komine-Kobayashi M, Tanaka R, Liu M, Mizuno Y, & Urabe T (2005) Edaravone reduces early accumulation of oxidative products and sequential inflammatory responses after transient focal ischemia in mice brain. *Stroke*, **36,** 2220-2225.
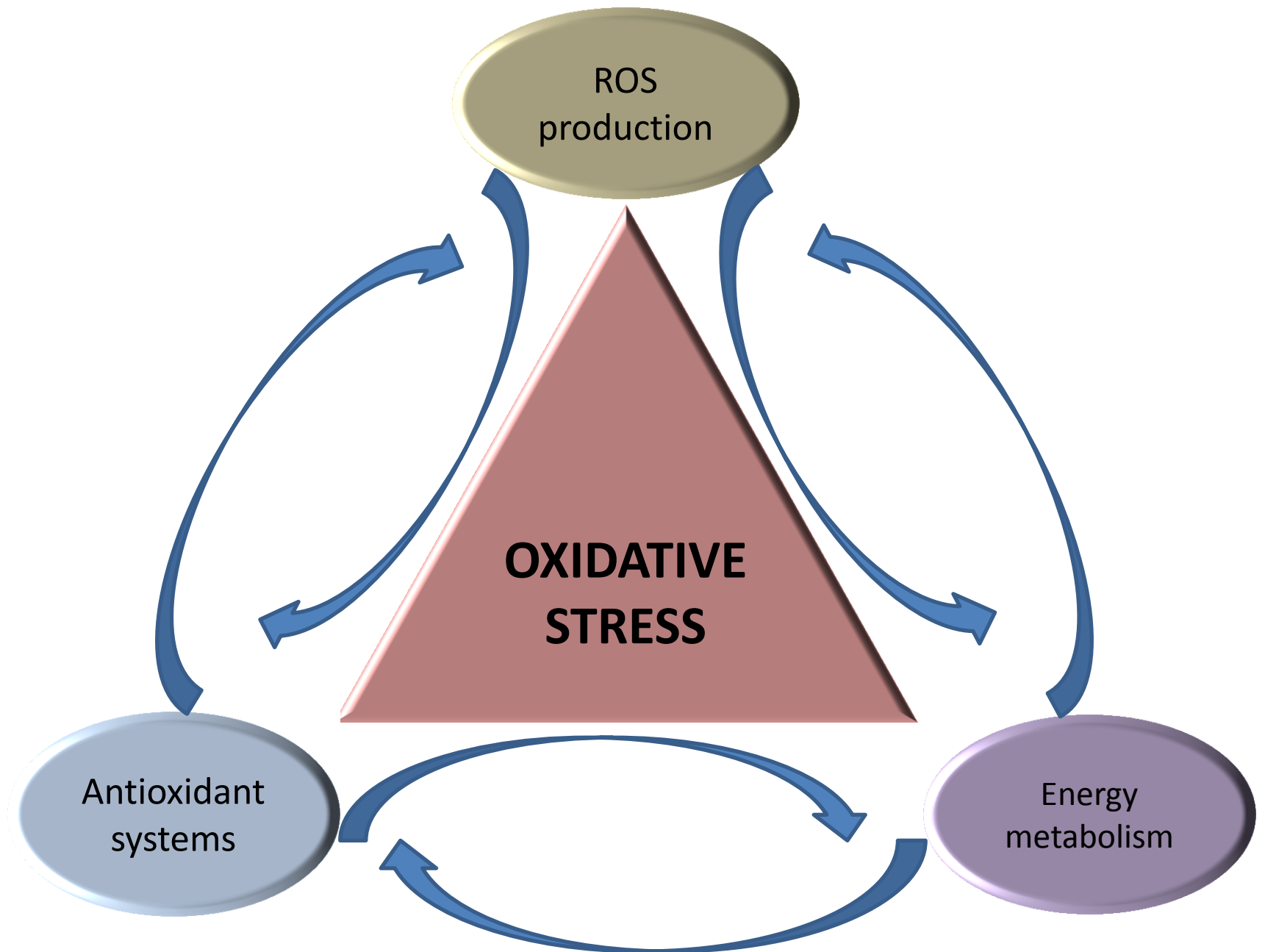
Figure 1

Figure 2